

# Analysis of *Prymnesium parvum* blooms in Lake Whitney, Texas

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# Overview

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- Problems:
  - Winter-Spring blooms of *Prymnesium parvum* in numerous Texas reservoirs (PK, Granbury, Whitney)
  - Frequent blooms of *P. parvum* in the upper Colorado River
  - Associated fish kills in reservoirs, rivers, and fish hatcheries

# Overview

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- Questions Identified by Texas HAB Team:
  - What factors control the development of *P. parvum* blooms in Texas reservoirs?
  - What physical, chemical, and biological characteristics trigger the build-up of *P. parvum* populations?
  - What causes toxin production in *P. parvum*?

# Overview

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- What factors contribute to *P. parvum* blooms?
  - Is there a relationship between environmental conditions and development of blooms?
  - Is this relationship mediated through algal population dynamics or trophic-level interactions?

# Approach - proposed studies

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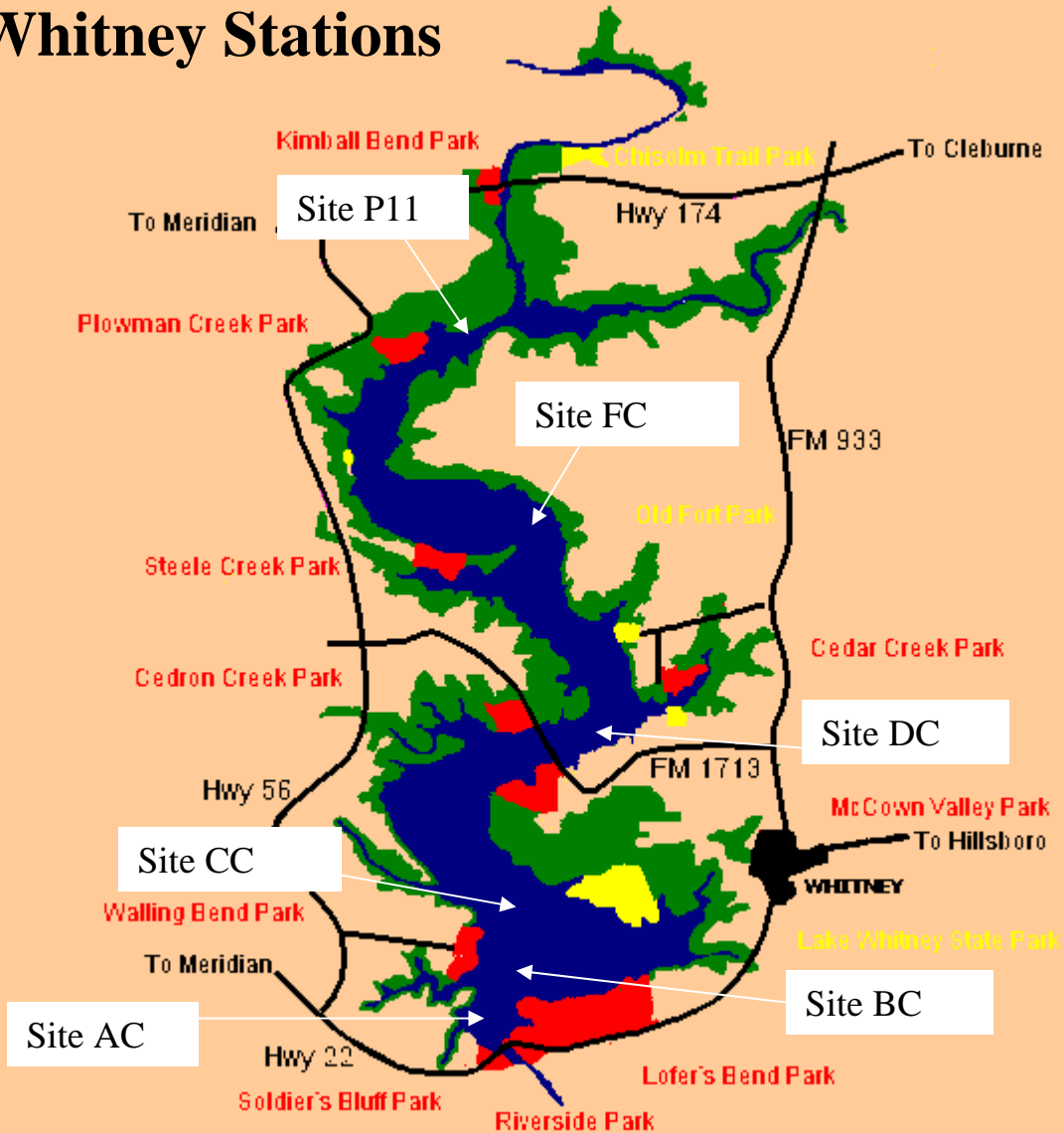
- Synoptic sampling of paired reservoirs during *P. parvum* blooms
  - Simultaneously assess population densities and the physical and chemical environmental gradients
  - Correlate biological responses to environmental gradients
- Experimental manipulation of important gradients
  - Nutrient enrichment experiments to ID potential limitation
  - Dilution bioassays to estimate importance of grazing
  - Functional response to limiting factors

# Pilot Study on Lake Whitney

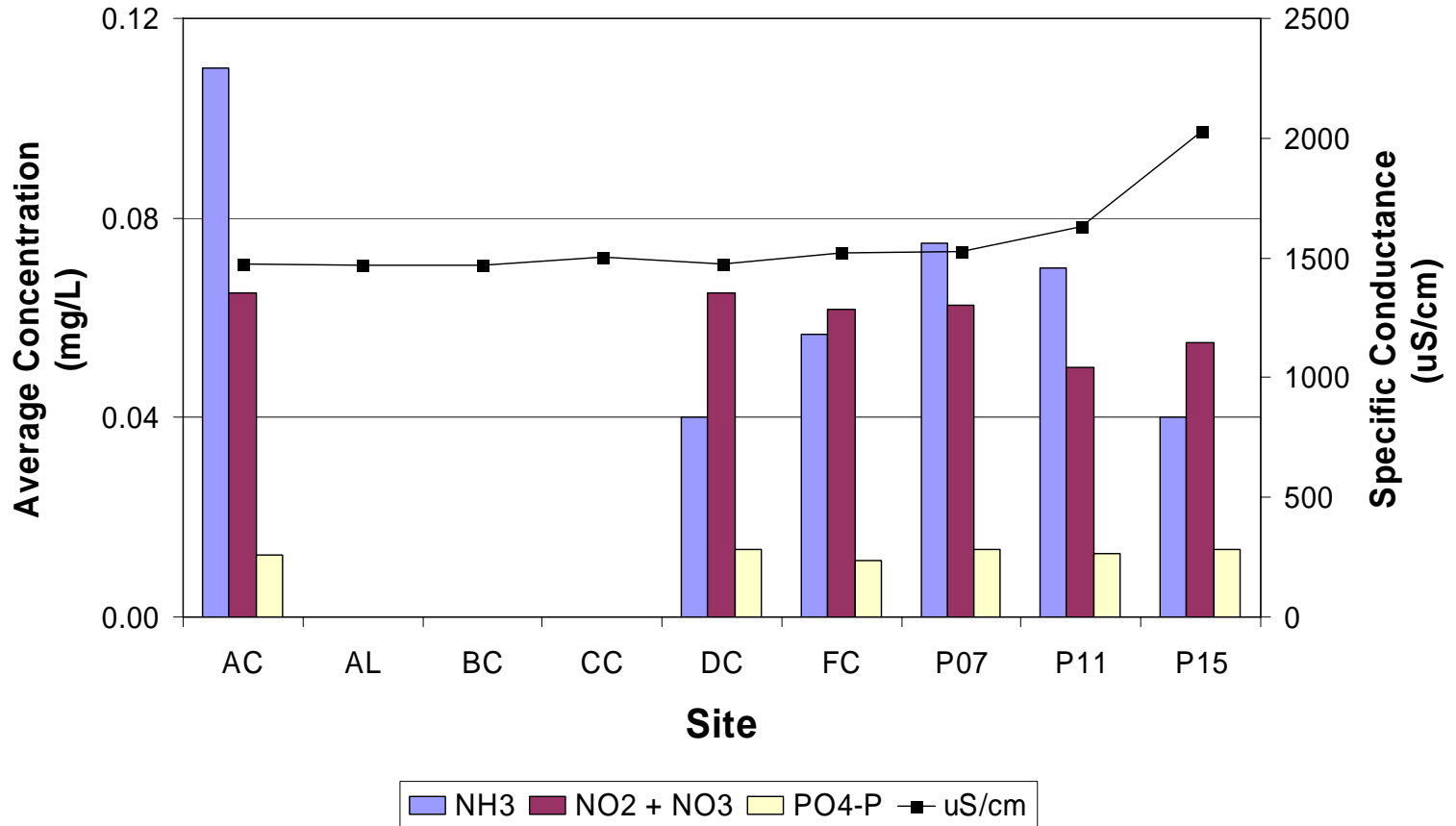
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- Assessment of biological responses to gradients in Lake Whitney
  - Synoptic cruises during the *P. parvum* bloom to document physical and chemical gradient
  - Experimental nutrient enrichment gradients to assess potential for nutrient limitation of *P. parvum* populations
  - Experimental grazing gradients to assess loss rates (dilution and addition)

# Lake Whitney Stations

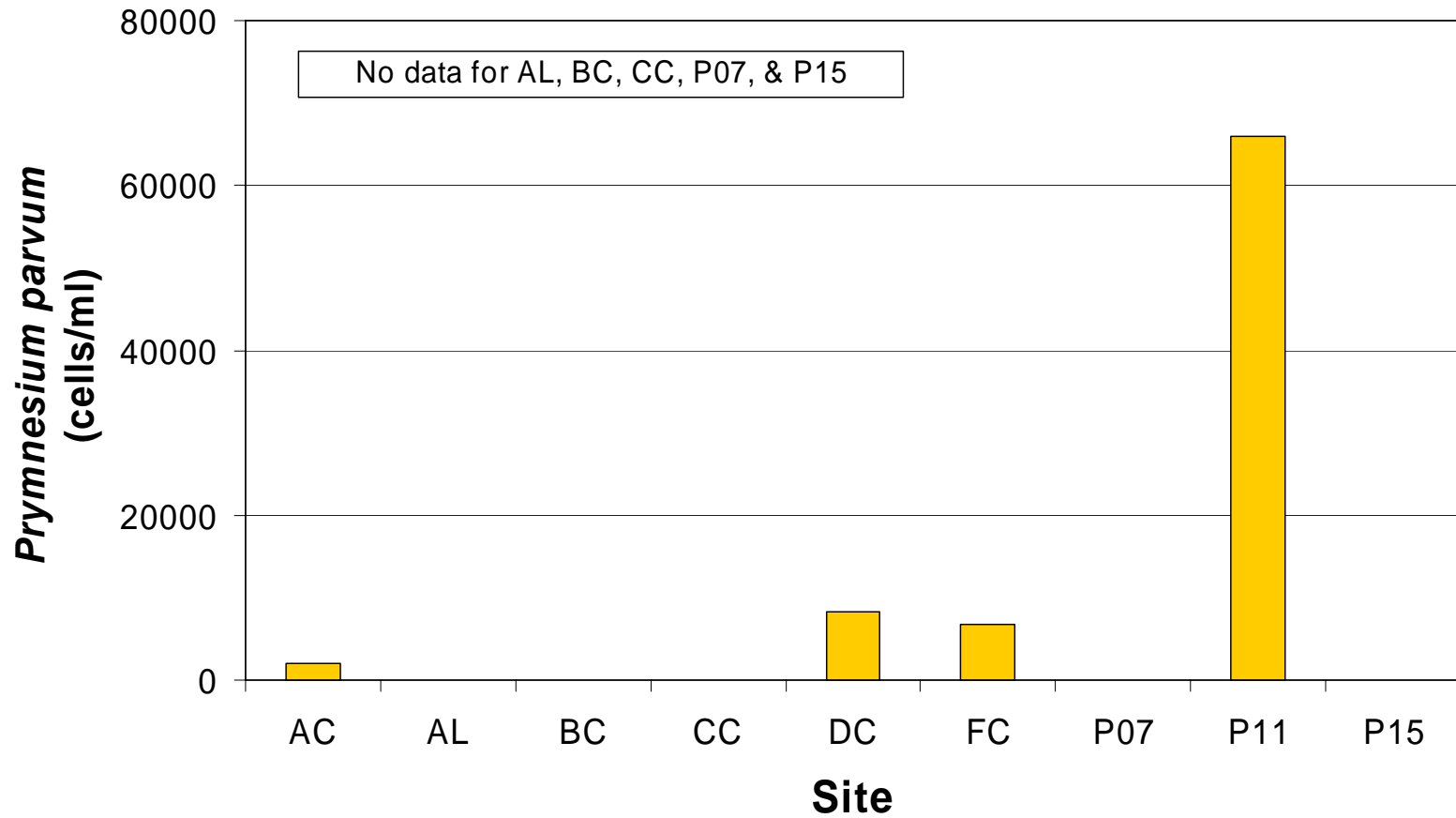


# Lake Whitney 2003 Ambient Chemistry: (all depths; Feb-April)





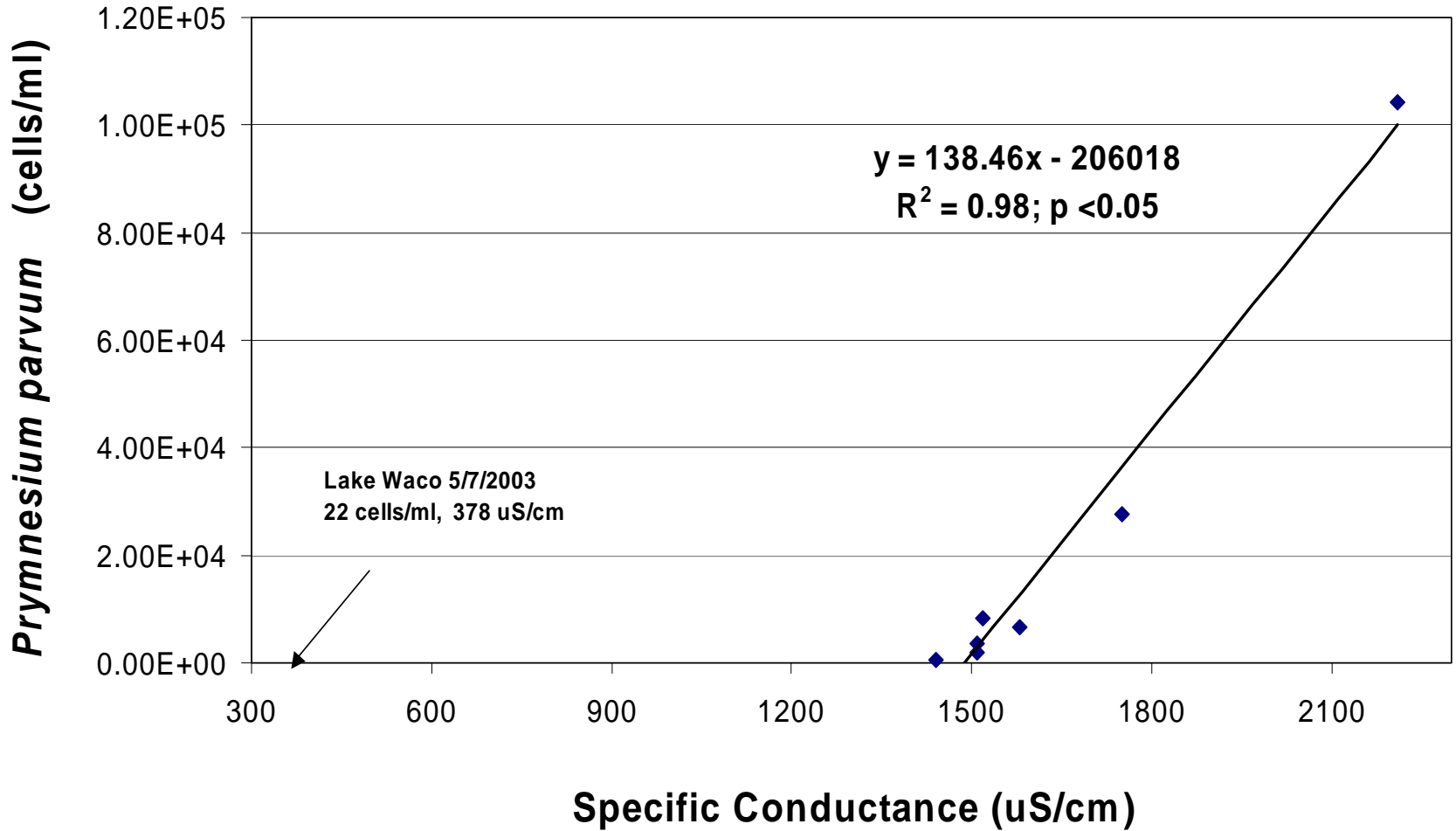
# Lake Whitney 2003 *P. parvum* abundance: (average for Feb-April)



# Correlations between *P. parvum* densities and selected variables

Lake Whitney <i>P. parvum</i> Surface Densities, Composite Nutrient Data					
	Cond.	NH <sub>3</sub> -N	NO <sub>2</sub> +NO <sub>3</sub>	PO <sub>4</sub> -P	CELLS/ML
Cond.	1.00	-0.49	-0.78	0.36	<b>0.98</b>
NH <sub>3</sub> -N		1.00	-0.01	-0.46	-0.51
NO <sub>2</sub> +NO <sub>3</sub>			1.00	0.16	<b>-0.72</b>
PO <sub>4</sub> -P				1.00	0.50
CELLS/ML					1.00

# Feb-April 2003 Lake Whitney: surface



Step-wise Multiple-Regression Summary: CELLS/ML  
 Surface Densities, Composite Nutrient Data

	Beta	Beta SE	B	B SE	t(1)	p-level
Intercpt			3907	13932	0.28	0.83
COND	0.52	0.03	73	4	18.53	0.03
NH <sub>3</sub> -N	-0.12	0.01	-113936	12177	-9.36	0.07
NO <sub>2</sub> +NO <sub>3</sub>	-0.37	0.02	-1869286	121535	-15.38	0.04
PO <sub>4</sub> -P	0.32	0.01	1876156	71168	26.36	0.02
R= .99 Adjusted R <sup>2</sup> = .99						
F(4,1)=5297.0 p<.01030 Std.Error of estimate: 615.29						

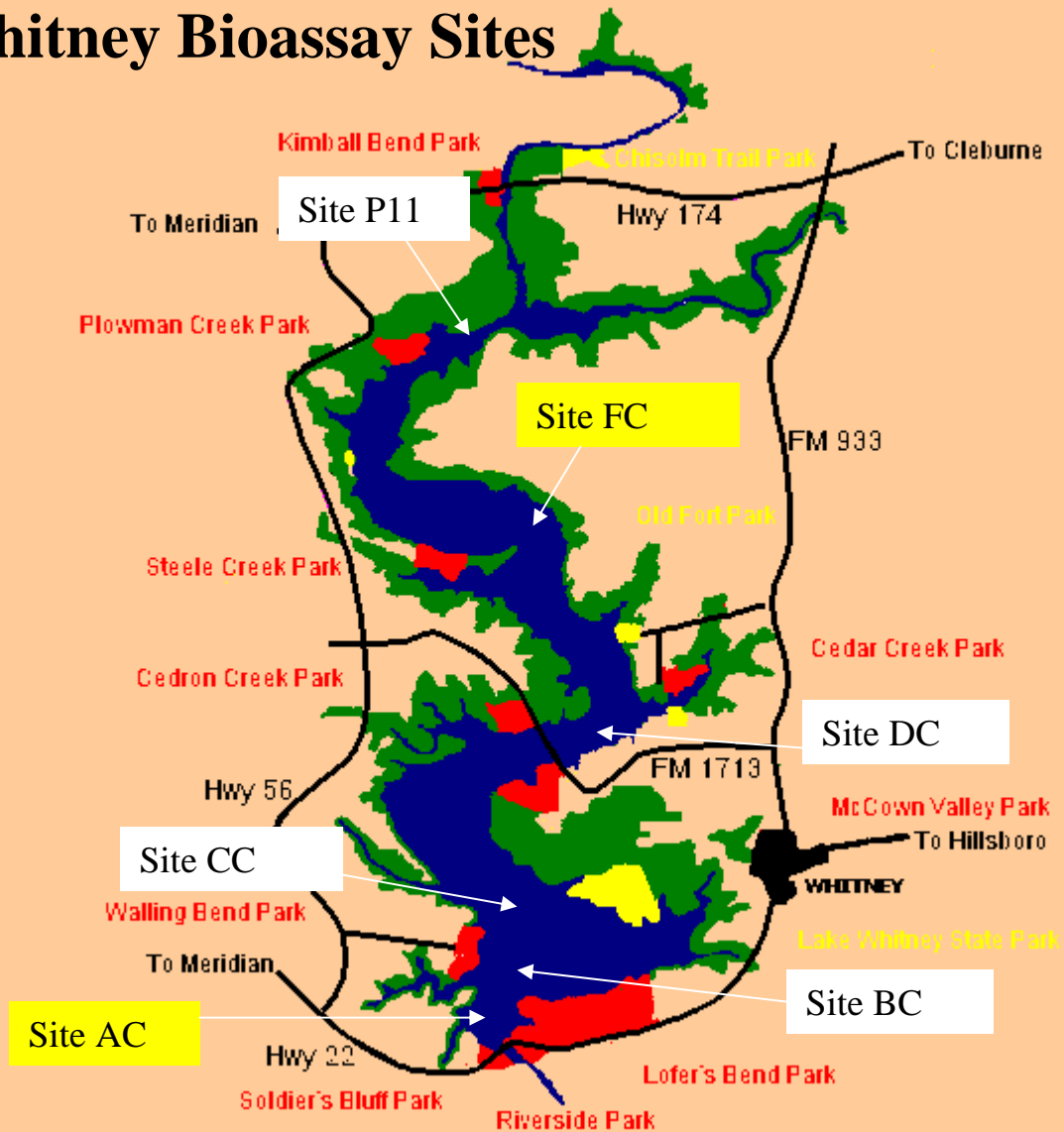
# Lake Bioassay Methods

- Acclimated growth-rate method using IVF followed by cell counts to estimate daily growth (  $r$  )
- Treatments included N, P, and Si additions to ambient lake water
- Laboratory incubations at ambient temperature and light lasted 8 days

# Lake Bioassay Methods (cont.)

- Five or six replicates per treatment
- Zooplankton removed using 153 $\mu$ m Nitex
- Results recorded using *in vivo* fluorescence
- Growth responses to treatments were calculated using an exponential growth model ( $N_t = N_o e^{rt}$ )
- Replicate estimates of (  $r$  ) for each treatment

# Lake Whitney Bioassay Sites

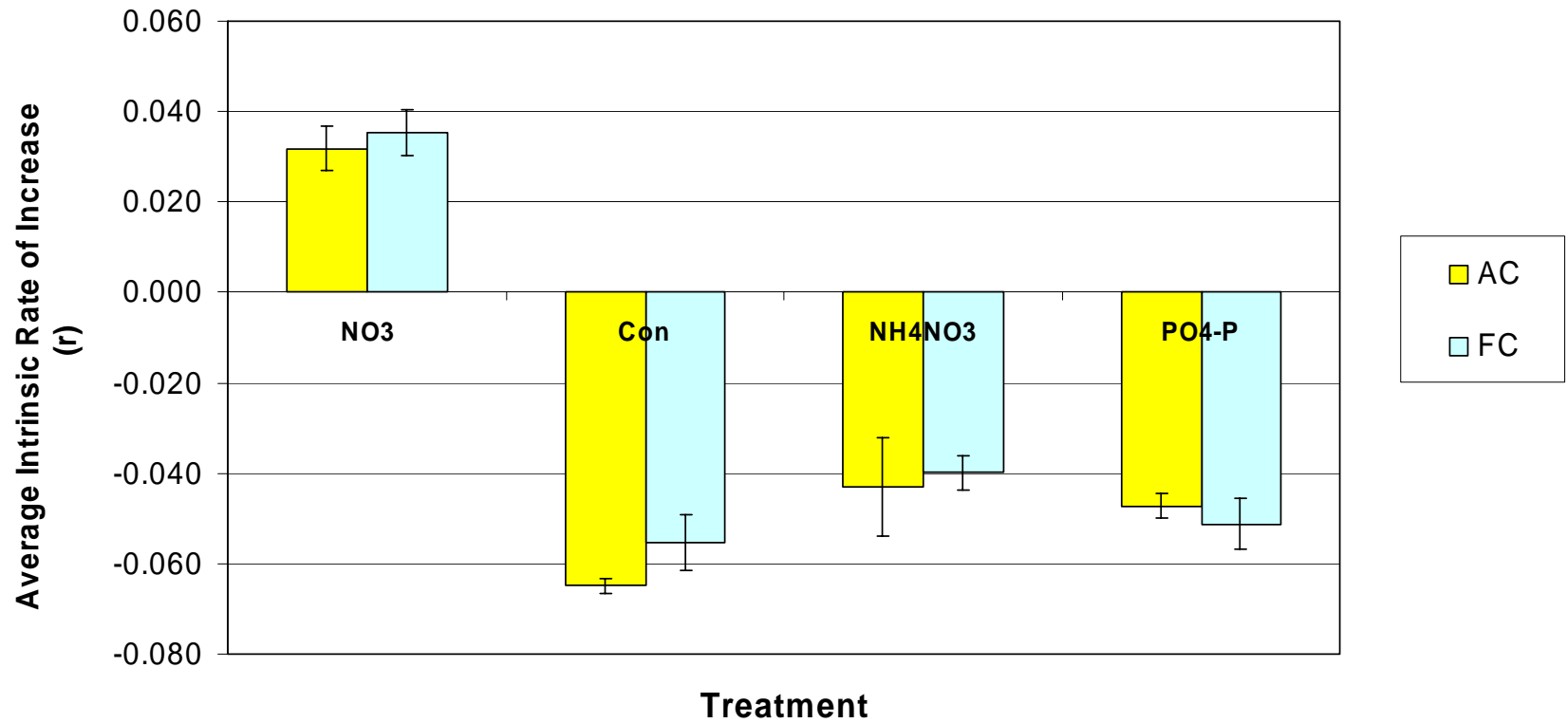


**Lake Whitney Nutrient Bioassay 4/29/03**

Site	Treatment	Mean r	SDEV	95% CI
AC	NO <sub>3</sub>	0.032	0.006	0.005
AC	Con	-0.065	0.002	0.002
AC	NH <sub>4</sub> NO <sub>3</sub>	-0.043	0.014	0.011
AC	PO <sub>4</sub> -P	-0.047	0.003	0.003
FC	NO <sub>3</sub>	0.035	0.006	0.005
FC	Con	-0.055	0.007	0.006
FC	NH <sub>4</sub> NO <sub>3</sub>	-0.040	0.004	0.004
FC	PO <sub>4</sub> -P	-0.051	0.007	0.006



## Lake Whitney Nutrient Bioassay 4/29/03



Lake Whitney Nutrient Bioassay 4/29/03:  
large volume, un-replicated design

Dilution Bioassay			
Site	Treatment	r	r <sub>uf</sub>
FC	50%D NO <sub>3</sub>	0.033	0.039
FC	50% D Con	-0.042	-0.037
FC	NO <sub>3</sub>	0.035	0.023
FC	Con	-0.055	-0.063
FC	NH <sub>4</sub> NO <sub>3</sub>	-0.040	
FC	PO <sub>4</sub> -P	-0.051	

# Original Questions

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- What factors contribute to *P. parvum* blooms?
  - Is there a relationship between environmental conditions and development of blooms?
  - Is this relationship mediated through algal population dynamics or trophic-level interactions?

## Results to date – April 2003 survey of Lake Whitney

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- *P. parvum* densities follow in-lake environmental gradient
- *P. parvum* growth is stimulated by nitrate addition during latter part of bloom
- *P. parvum* net growth rates may be sensitive to species interactions

## Next steps?

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- Continue paired-reservoir assessment between Lake Whitney and Lake Waco - add others
- Continue to assess importance of grazing and nutrient limitation for *P. parvum* populations
- Assess toxin levels as a function of *P. parvum* density, conductivity, and nutrient limitation
- Explore options for Lake Whitney sediment core analysis using *P. parvum* biomarkers

# Acknowledgements

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- Texas HAB Team, TPWD
- Joan Glass, TPWD
- Dan Roelke, TAMU
- US Army Corps of Engineers
- USGS
- Texas Commission on Environmental Quality

# DY III Media Stock Additions

Nutrient	Final Concentration
$\text{Na}_2\text{HPO}_4$	8 mg/L
$\text{NH}_4\text{NO}_3$	5 mg/L
$\text{NaNO}_3$	20mg/L
$\text{Na}_2\text{SiO}_3\cdot 9\text{H}_2\text{O}$	30 mg/L